

1 **Rapid Detection of Carbapenemases in *Enterobacteriaceae*: Evaluation of the**
2 **RESIST-3 O.K.N (OXA-48, KPC, NDM) Multiplexed Lateral Flow Assay**

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25 Running title: Multiplexed CRE lateral flow cards

26 **Text**

27 The identification, treatment and control of carbapenem resistant *Enterobacteriaceae*
28 (CRE) infections are a major challenge for healthcare institutions and diagnostic
29 laboratories worldwide. Those producing plasmid mediated OXA-48, KPC, NDM or
30 VIM-like carbapenemases (CPE) are most concerning, being frequently involved in
31 nosocomial outbreaks that are difficult and very costly to manage (1). A simple and
32 rapid test, able to provide sensitive and specific identification of CRE and also the
33 carbapenemase present, is critical in any strategy aimed at addressing this problem.

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35 Numerous phenotypic (MIC determination, disc diffusion, selective culture media,
36 acidometric) and genotypic (PCR amplification, microarrays, DNA sequencing)
37 methods have been used in the laboratory isolation, diagnosis and confirmation of
38 CRE (2). Recently, two novel immunochromatographic lateral flow assays (Coris
39 BioConcepts, Gembloux, Belgium) were developed for the specific detection of OXA-
40 48 (OXA-48 K-SeT) and KPC-like carbapenemase producers (KPC K-SeT). These
41 assays have been evaluated in multiple laboratories, using diverse sets of organisms
42 (*Escherichia coli*, *Klebsiella*, *Enterobacter*, *Serratia*, *Providencia*, *Pseudomonas* spp)
43 carrying multiple β -lactamase, OXA-48 (OXA-48, 162, 181, 204, 232, 242) and KPC
44 (KPC-2/3/4) allelic variants. (3,4,5,6). Both have a reported sensitivity and specificity
45 of 100 % when compared to molecular detection of carbapenemase genes as the
46 gold standard (3,4,5,6). They have also been shown to be compatible with
47 organisms recovered from most un-supplemented, selective, solid and liquid culture
48 media currently in use in diagnostic laboratories and with bacteria taken directly from
49 positive blood culture bottles (BacT/ALERT, bioMerieux, Macy L'Etoile, France) or
50 culture positive urinary samples (3, 4). A modification of this system - RESIST-3

51 O.K.N (Coris BioConcepts) designed for the simultaneous detection of OXA-48, KPC
52 and NDM-like enzymes using a single disposable cartridge, has now been
53 manufactured. Here we assessed the ability of the RESIST-3 O.K.N assay to detect
54 OXA-48, KPC and NDM co-production in a collection of 112 non-replicate well
55 defined CRE isolates received in our laboratories.

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57 All isolates were carbapenem resistant *Enterobacteriaceae* (ertapenem MIC >
58 1µg/ml) recovered from routine clinical samples submitted to Barts Health NHS Trust
59 or the Antimicrobial Research Laboratory at Queen Mary University London
60 between 1/2011 and 12/2016. Identification was performed using the Bruker MALDI-
61 ToF mass spectrometry system (Bruker UK Ltd, Coventry, UK), with carbapenem
62 MICs determined using MicroScan WalkAway Negative Combo 36 panels (Siemens
63 Healthcare Diagnostics, Deerfield, IL) and confirmed with Etest gradient strips
64 (bioMerieux). Multiplex PCRs were used to confirm the presence of class A (KPC,
65 BKC, SME, VEB, PER, GES), B (IMP, VIM, NDM, SIM, SPM, DIM, GIM, KHM, FIM,
66 AIM, DIM, TMB, FRI), and D (OXA-like) carbapenemase genes and all allelic
67 variants identified were confirmed by Sanger sequencing of the entire coding regions
68 (7, 8). The collection consisted of carbapenemase producing *K. pneumoniae* (n=57),
69 *E. coli* (n=17), *Enterobacter spp* (n=14), *Providencia stuartii* (n=3) and single isolates
70 of *Citrobacter koseri*, *Morganella morganii* and *Proteus mirabilis* (Table 1).
71 Carbapenemases produced by the isolates were identified as KPC-2/4, NDM-1/5/7,
72 VIM-1/4, IMP-1, OXA-48 and OXA-232. Nine isolates produced 2 carbapenemases,
73 either OXA-48 in combination with NDM-1 (n=6) or KPC-2 with VIM-1 (n=3).

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75 Detection of OXA-48, KPC and NDM using the RESIST-3 O.K.N. cassettes was
76 carried out according to the manufacturer's protocol. Bacteria were grown for 18 hrs
77 at 37°C on Mueller-Hinton II agar plates (Oxoid) and a single colony emulsified in 5
78 drops of the lysis buffer. Cassettes were loaded with 3 drops of lysate and read
79 within 5 minutes. Six carbapenem susceptible Gram-negative type strains (*K.*
80 *pneumoniae* NCTC 9633, *E. coli* NCTC 12241, *E. cloacae* NCTC 13380, *E.*
81 *aerogenes* NCTC 9375, *P. aeruginosa* ATCC 27852, *Acinetobacter baumannii*
82 ATCC 19606) were used as negative controls. Eighteen additional
83 *Enterobacteriaceae* isolates [*K. pneumoniae* (8) / *oxytoca* (1), *E. coli* (4), *E. cloacae*
84 (2) / *aerogenes* (2), *S. marcesens* (1)] with phenotypic resistance to ertapenem but
85 without a known carbapenemase were also used as negative controls.

86

87 There was complete agreement between the carbapenemases detected by PCR and
88 the results obtained with RESIST-3 O.K.N (Table 1). The lateral flow device was able
89 to correctly identify and differentiate OXA, KPC and NDM production amongst all
90 CRE species and enzyme variants tested, including those carrying >1
91 carbapenemase. No cross reactions were observed for strains with carbapenem
92 resistance due to IMP-1 or VIM-1/4 (Table 1), with any of the susceptible type strains
93 and, with the 18 CRE that had no OXA, KPC or NDM carbapenemase detectable by
94 PCR (Figure 1).

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96 We found the RESIST-3 O.K.N assay to be 100 % sensitive and specific in the
97 detection and differentiation of OXA-48, KPC and NDM-like carbapenemases
98 amongst CRE recently referred to our laboratory. Although the assay does not
99 currently extend to the detection of VIM or IMP-like metallo- β -lactamases, the ability

100 to detect 3 of the 5 most prevalent and transmissible carbapenemases found
101 worldwide is a significant advantage. The ease of use, speed and low cost (< \$15) of
102 the cassettes make it attractive as a diagnostic tool for use in the management of
103 CRE outbreaks. It's role either as a primary screening, confirmatory or rapid point-of
104 care test should be assessed further, ideally prospectively in the setting of a
105 polyclonal nosocomial CRE outbreak.

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107 **Acknowledgements**

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109 Bioconnections (Knypersley, UK). No other specific funding was used to undertake
110 this study as it was performed as part of our routine activities. Both authors declare
111 no conflict of interest and have no association with CORIS BioConcept.

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118 **Table 1:** Detection of Carbapenemase Production using the RESIST-3 O.K.N assay

Isolate	Carbapenemase Carried	RESIST-3 O.K.N Result		
		OXA-48-like (O)	KPC-like (K)	NDM-like (N)
<i>Klebsiella pneumoniae</i>	OXA-48 (23)	23	0	0
	OXA-48 + NDM-1 (4)	4	0	4
	OXA-232 (6)	6	0	0
	KPC-2 (8)	0	8	0
	KPC-2 + VIM-1 (1)	0	1	0
	NDM-1 (14)	0	0	14
	VIM-4 (1)	0	0	0
<i>Escherichia coli</i>	OXA-48 (1)	1	0	0
	KPC-2 (3)	0	3	0
	KPC-2 + VIM-1 (1)	0	1	0
	NDM-1 (6)	0	0	6
	NDM-5 (3)	0	0	3
	NDM-7 (2)	0	0	2
	VIM-1 (1)	0	0	0
<i>Enterobacter cloacae</i>	OXA-48 + NDM-1 (2)	2	0	2
	NDM-1 (2)	0	0	2
	KPC-4 (2)	0	2	0
	IMP-1 (1)	0	0	0
	VIM-1 (4)	0	0	0
<i>Enterobacter aerogenes</i>	KPC-2 (2)	0	2	0
	NDM-1 (1)	0	0	1
<i>Citrobacter koseri</i>	KPC-2 + VIM-1 (1)	0	1	0
<i>Providencia stuartii</i>	NDM-1 (1)	0	0	1
	VIM-1 (2)	0	0	0
<i>Morganella morganii</i>	VIM-1 (1)	0	0	0
<i>Proteus mirabilis</i>	VIM-1 (1)	0	0	0

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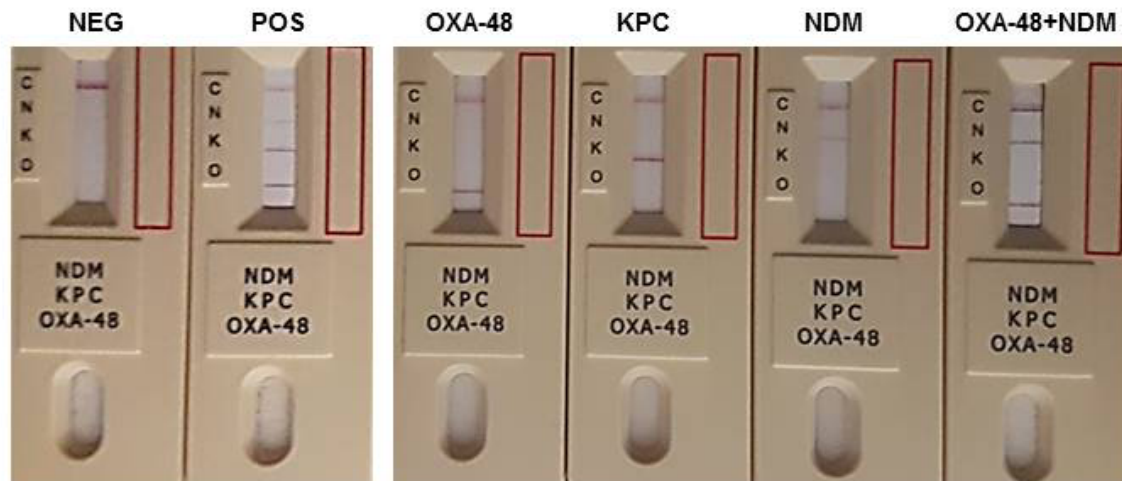
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124 **Figure 1:** Detection of OXA-48, KPC and NDM-like carbapenemase in isolates producing single and dual enzymes using the
125 RESIST-3 O.K.N assay



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129 **References**

- 130 1. **Otter JA, Burgess P, Davies F, Mookerjee S, Singleton J, Gilchrist**
131 **M, Parsons D, Brannigan ET, Robotham J, Holmes AH.** 2016. Counting the cost
132 of an outbreak of carbapenemase-producing Enterobacteriaceae: an economic
133 evaluation from a hospital perspective. *Clin Microbiol Infect* S1198-743X(16)30464-5.
134 doi: 10.1016/j.cmi.2016.10.005
- 135
- 136 2. **Lutgring JD, Limbago BM.** 2016. The Problem of Carbapenemase-Producing-
137 Carbapenem-Resistant-Enterobacteriaceae Detection. *J Clin Microbiol* 54 (3):529-34
- 138
- 139 3. **Wareham DW, Shah R, Betts JW, Phee LM, Momin MH.** 2016. Evaluation of an
140 Immunochromatographic Lateral Flow Assay (OXA-48 K-SeT) for Rapid Detection of
141 OXA-48-Like Carbapenemases in Enterobacteriaceae. *J Clin Microbiol.* 54(2):471-3.
142 doi: 10.1128/JCM.02900-15.
- 143
- 144 4. **Glupczynski Y, Evrard S, Ote I et al.** 2016. Evaluation of two new commercial
145 immunochromatographic assays for the rapid detection of OXA-48 and KPC
146 carbapenemases from cultured bacteria. *J Antimicrob Chemother*; 71: 1217–22.
- 147
- 148
- 149 5. **Dortet L, Jousset A, Sainte-Rose V, Cuzon G, Naas T.** 2016. Prospective
150 evaluation of the OXA-48 K-SeT assay, an immunochromatographic test for the
151 rapid detection of OXA-48-type carbapenemases. *J Antimicrob*
152 *Chemother.* 71:1834-40. doi: 10.1093/jac/dkw058.

153

154 6. Meunier D, Vickers A, Pike R, Hill RL, Woodford N, Hopkins KL. 2016.

155 Evaluation of the K-SeT R.E.S.I.S.T. immunochromatographic assay for the rapid
156 detection of KPC and OXA-48-like carbapenemases. J Antimicrob Chemother
157 71:2357-9. doi: 10.1093/jac/dkw113.

158

159 7. Oikonomou O, Liakopoulos A, Phee LM, Betts J, Mevius D, Wareham DW.

160 2016. Providencia stuartii Isolates from Greece: Co-Carriage of Cephalosporin (*bla*SHV-5,
161 *bla*VEB-1), Carbapenem (*bla*VIM-1), and Aminoglycoside (*rmtB*) Resistance Determinants
162 by a Multidrug-Resistant Outbreak Clone. Microb Drug Resist. 22:379-86. doi:
163 10.1089/mdr.2015.0215.

164

165 8. Dallene C, Da Costa A, Decre D, Favier C, Arlet G. 2010. Development of a set of
166 multiplex PCR assays for the detection of genes encoding β -lactamases in
167 Enterobacteriaceae. J Antimicrob Chemother. 65: 490-495

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